

Generation Using a Three Dimensional Hepatic Matrix

erry^{2,4}, Li Zhang^{1,4}, Junji Komori^{3,4}, Eric Lagasse^{3,4}, Szymanski^{1,2,4,*}

¹Regenerative Medicine, University of Pittsburgh, Pittsburgh PA
²Biomedical Engineering, University of Pittsburgh, Pittsburgh PA
³Cell Biology, University of Pittsburgh, Pittsburgh PA
⁴Center for Regenerative Medicine, University of Pittsburgh,

(s.edu)

options for the treatment of end stage liver disease. The current standard is liver transplantation but organ donor shortage is a major barrier with little hope of improvement in the foreseeable future. Hepatic scaffolds composed of extracellular matrix (ECM) provide a promising platform for the constructive remodeling of organs such as liver, bladder, and skin. The present study involves the reconstruction of whole rat livers by retrograde perfusion. The remaining ECM matrix, when continuously perfused, retains its three-dimensional structure, including microvasculature. Seeding of the scaffold with primary hepatocytes has shown maintenance of cell differentiation and the retention of hepatocellular function by albumin production, EROD metabolism and ammonia production. Hepatic ECM scaffolds were recellularized using three different methods in an effort to determine which method was optimal. Sheets of primary hepatocytes with morphologically normal appearing cell to cell junctions were observed following recellularization and in-vitro culture. These scaffolds constitute functional three-dimensional hepatic parenchyma and may be used for regenerative medicine therapy for patients with end stage

416 Expression of Liver Specific Functions in Rat Hepatocyte Spheroid Array

Yusuke Sakai, Seita Yamagami, and Kohji Nakazawa*

Department of Life and Environment Engineering, The University of Kitakyushu, 1-1 Hibikino, Wakamatsu-ku, Kitakyushu, Fukuoka 808-0135, Japan

(*nakazawa@env.kitakyu-u.ac.jp)

It is known that spherical multicellular aggregates (spheroids), which are formed by the rearrangement and compaction of a cell aggregates, have a tissue-like structure and can express higher levels of liver-specific functions as compared to the traditional hepatocyte monolayer. In this study, we developed a spheroid array system and evaluated the expression of genes encoding key molecules involving in liver-specific functions, namely, cell adhesion molecules, transcription factors, protein and metabolic enzymes, and transporters.

The hepatocyte spheroid array chip was manufactured by combination of the microfabrication and the microcontact printing. The chip comprised 672 circular microwells in a triangular arrangement; each well was 300 µm in diameter, 200 µm in depth, and 400 µm in pitch. The center of each microwell had a 100-µm cell adhesion area was modified with collagen, and the remaining part of each microwell was modified with polyethylene glycol to create the non-adhesive area. The hepatocytes cultured on the chip gradually aggregated on the collagen-coated area in each microwell and had developed into a smooth spheroid had a uniform diameter. Though the expression levels of all the genes encoding key molecules in the spheroids tended to decrease gradually with culture time, they were consistently higher than those in the monolayer culture for at least 10 days of culture.

These results suggest that hepatocyte spheroids acquire intercellular organization and maintain many metabolic functions. Thus, this hepatocyte spheroid array system seems to be a promising model for various *in vitro* cell-based assays.

Three-Dimensional Primary Hepatocyte Culture on Heparin-based Hydrogel

Junghyun Lee,² Caroline Jones,² Alexander Revzin,^{2,*} and

¹Department of Biomolecular Nanotechnology and Department of Chemical and Engineering, Gwangju Institute of Science and Technology, Gwangju, Korea

²Biomedical Engineering, University of California, Davis,

(s.edu, **gytae@gist.ac.kr)

Hepatocytes, a main cell type in liver, carry out most of the functions of liver, including production of majority of the plasma proteins, regulation of the carbohydrate, urea, and lipid metabolism as well as detoxification of exogenous chemicals. However, primary hepatocytes rapidly lose their functions and viability after transplantation. This study is focused on the maintenance of their function for a long-term in *in vitro* primary hepatocyte culture. Formation of hepatocytes in 3-D is one of the approaches for a long-term culture. The use of hydrogels is a promising way because of their mechanical strength, and the easy tuning of their modulus to that of the native tissue. In this study, we developed an in situ forming heparin-based hydrogel in the presence of biomolecules, such as cells. The hydrogel was formed by a Michael-type addition reaction between heparin and diacrylated poly (ethylene glycol). Heparin is interacting with numerous ECM proteins and growth factors through its heparin-binding domain. Particularly, heparin is one of the major components in the matrix of liver, and it has a high affinity for hepatocyte growth factor (HGF), well known as a potent factor in growth and differentiation of hepatocytes. In this study, we applied the heparin-based hydrogel to primary hepatocyte culture. The initial viability of hepatocytes on the hydrogel was sufficiently high (over 70%), and the long-term viability as well as functions over several weeks were maintained. The effects of adding HGF, using spheroids of hepatocytes, and applying co-culture with stellate cell also will be

418 Mesenchymal Stem Cell Therapy Ameliorates Acute Liver Injury in Rabbits

Hsiao-huei Chao,^{1,*} Tsung-Min Lin,² Hui-Ying Chung², Long-bin Jeng³ and Sophia Chia-Ning Chang⁴

¹Department of Pediatric Surgery, ²Department of Medical Research, ³Department of General Surgery, ⁴Department of Plastic Surgery, China Medical University Hospital, Taichung 404, Taiwan

(*mimiwhere@hotmail.com)

Acute liver failure is characterized by rapid deterioration of liver function and a high mortality. Currently, there are no specific therapies except liver transplantation. However, shortage of donor organs remains the major obstacles. Cell therapy of acute liver failure should provide rapid support for the injured liver. However, adult human hepatocytes have a poor proliferative potential. Bone marrow comprising heterogeneous cell populations contains certain progenitors with the ability to differentiate into multiple mesenchymal cell lineages. To identify any differentiation plasticity of adult bone marrow mesenchymal stem cells (MSCs), we used allyl alcohol (AA) treated rabbits for an acute liver failure model.

The MSCs were aspirated and expanded to a cellular concentration of $1 \times 10^8/4$ ml. The rabbits were injected AA intraperitoneally. Three days after the injection, the MSCs were injected via portal vein. The control group rabbits were injected with 4 ml of normal saline under the same condition. The postoperative recovery was closely monitored by hepatic function markers. Decreased portal fibrosis after stem cell injection was noted by Masson's trichrome stain, while the reticulin framework restoration was also vaguely identifiable by reticulin stain.

Whether extracorporeal devices or the transplantation of primary hepatocytes, stem cells or cells genetically engineered to over-express key metabolic functions, a proliferative phenotype or cytoprotective pathways will be best suited to meeting these demanding challenges.